

Opinion

Synaptic Tenacity or Lack Thereof:
Spontaneous Remodeling of SynapsesNoam E. Ziv^{1,2,*} and Naama Brenner^{1,3}

Synaptic plasticity – the directed modulation of synaptic connections by specific activity histories or physiological signals – is believed to be a major mechanism for the modification of neuronal network function. This belief, however, has a ‘flip side’: the supposition that synapses do not change spontaneously in manners unrelated to such signals. Contrary to this supposition, recent studies reveal that synapses do change spontaneously, and to a fairly large extent. Here we review experimental results on spontaneous synaptic remodeling, its relative contributions to total synaptic remodeling, its statistical characteristics, and its physiological importance. We also address challenges it poses and avenues it opens for future experimental and theoretical research.

Synaptic Plasticity and Synaptic Tenacity

At a conceptual level, the central nervous system (CNS) comprises a vast network of excitable cells connected by specialized junctions known as synapses. Long before excitability and synaptic transmission were fully understood, scholars such as William James [1] suggested that changes to connections (‘tracts of conduction’), driven by sequential or concomitant activation of ‘elementary brain-processes’, might explain the formation of new associations and the learning of new tasks. In fact, James credits this idea to yet earlier scholars (e.g., Descartes, Locke) and notes that they ‘hit upon this explanation, which modern science has not yet succeeded in improving’ [1]. Years later, Donald Hebb rephrased this idea, using it as a cornerstone for constructs such as cell assemblies, thereafter used to explain network function and ultimately behavior in physiological terms [2]. Inspired by these theories, and armed with increasingly powerful methodologies, numerous investigators have convincingly demonstrated that **activity histories** (see [Glossary](#)) can influence and change such tracts of conduction, now largely identified as chemical synapses. The resulting changes, generally referred to as synaptic plasticity, are thus commonly viewed as evidence for this venerable theory.

The long-held framework described above has an implicit yet often ignored corollary, which is the supposition that synapses, when not driven to change by particular activity patterns or histories, retain their properties indefinitely. This supposition would seem to be essential because without it, spontaneously occurring changes might drive spurious modifications in network function or undo physiologically relevant ones. Thus, to fully appreciate the importance and limitations of synaptic plasticity, it is essential to measure and understand the capacity of synapses to maintain their particular properties (e.g., probability of release, total receptor conductance, size, morphology, ultrastructure, composition) over behaviorally relevant time-scales. We refer to this capacity as **synaptic tenacity** [3–6].

The aim of this Opinion is to discuss challenges to the notion of synaptic tenacity that come from general biological considerations and experimental findings. Such findings collectively suggest that synaptic tenacity is inherently limited, since synapses do change spontaneously

Highlights

Synaptic remodeling is driven by both activity-dependent and spontaneous processes.

The magnitude of spontaneous synaptic remodeling is comparable with that of activity-dependent remodeling.

Spontaneous synaptic remodeling processes can give rise to a full repertoire of synaptic sizes even in the complete absence of activity.

While spontaneous remodeling processes drive continuous fluctuations in the properties of individual synapses, stable and skewed distributions of the same properties emerge as population invariants.

Scaling of synaptic size distributions appears as a population-level phenomenon when spontaneous remodeling processes are modulated by perturbations of spontaneous network activity.

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and to a fairly large extent. We survey these findings and discuss their implications in light of the supposition mentioned above. We then present principles emerging from recent literature on the manners by which spontaneous processes might govern dynamic and statistical properties of synaptic populations. We end by formulating several key questions concerning relationships between **spontaneous remodeling**, underlying biological processes, neuronal activity, and network function.

Synaptic Tenacity: Inherent Challenges

CNS synapses are micrometer-size, intricate assemblies of diverse molecules (e.g., receptors, ion channels, synaptic vesicle, scaffolding, cytoskeletal, adhesion, and signaling molecules). Typical lifetimes of synaptic molecules (i.e., days; see [7]) are orders of magnitude shorter than the lifetimes of many, if not most CNS synapses (weeks and months, and in some species probably years [8–10]; see [11] for a recent review). Consequently, the maintenance of synaptic connections involves continuous removal and degradation of molecules and their substitution with freshly synthesized copies. Given that CNS synapses are often remote from the neuron's somatic biosynthetic machinery (residing at distances of many centimeters or even meters, in some organisms), long-term synaptic maintenance is a remarkable biological feat [12,13]. Importantly, however, given these logistic challenges and the low copy numbers of many molecules at individual synapses, it is probably unrealistic to expect that synapses maintain their particular contents and, by extension, their functional properties with pinpoint precision (see [14]). This expectation is further challenged by the fact that synapses are not rigid structures but rather are dynamic assemblies of molecules (and organelles) that continuously migrate into, out of, and between neighboring assemblies through lateral diffusion, active trafficking, endocytosis, and exocytosis [15,16]. These molecular dynamics are often further accelerated by cellular processes associated with synaptic transmission, such as membrane recycling and cytoskeletal dynamics.

Conceivably, challenges to synaptic tenacity posed by continuous molecular turnover and the myriad molecular dynamics might be met somehow, resulting in roughly stable synaptic properties. More likely, however, molecular dynamics coupled with imprecise proteostasis will drive spontaneous changes in synaptic properties that have little to do with specific activity histories [17]. As we show next, experimental evidence seems to favor the latter possibility.

Spontaneous Synaptic Remodeling in Developing Networks

During development the brain is strongly influenced by experience and activity, yet it seems that activity is largely dispensable for synapse formation *per se*. Although this conclusion is well established [18,19] (see [20] for a review of earlier work), recent studies have highlighted observations that are particularly pertinent to this Opinion.

Studies in primary culture have shown that general properties of excitatory and inhibitory synapses are largely preserved even when networks develop in the complete absence of activity. For example, chronic suppression of neurotransmitter release or network activity in primary cultures of hippocampal neurons does not appear to affect synaptic spatial densities, colocalization of pre- and postsynaptic molecules, or the synaptic contents of important **postsynaptic density (PSD)** proteins (e.g., PSD-95/Dlg4, CaMKII α , SynGAP, Gephyrin) and neurotransmitter receptor subunits (Gria2/3, GABA α 2) [18,19]. Recently, these findings were confirmed and extended to additional preparations, including the intact brain. In one study [21] (see also [22]), spine types, spine spatial densities, and synaptic currents were examined in hippocampal organotypic cultures prepared from mice in which presynaptic release was

Glossary

Active zone (AZ): a spatially confined, electron-dense specialization of the presynaptic membrane; comprises specific proteins that orchestrate synaptic vesicle exocytosis and confine it to the presynaptic site.

Activity history: the history of synaptic activation through stimuli from natural sources (e.g., sensory input, internal activity) or through direct experimental stimulation in terms of the frequencies, durations, and temporal patterns of action potentials occurring in presynaptic (and postsynaptic) neurons or their respective compartments (axons, boutons, dendrites, and spines).

Distribution scaling: the collapse of two distributions into one curve following the scaling (stretching or compressing) of their x-axis. This property implies that the two distributions have the same shape but a different scale. Scaling can, in principle, result from the multiplication of all individual elements in the population by a fixed constant; it can, however, also result from non-uniform transformations.

Postsynaptic density (PSD): a spatially confined, electron-dense specialization of the postsynaptic membrane comprising numerous scaffolding, cytoskeletal, and signaling molecules; concentrates neurotransmitter receptors such that they are juxtaposed against presynaptic neurotransmitter release sites.

Skewed distribution: a distribution that is not symmetric around its mean (as is the Gaussian distribution) but rather skewed toward one side, which has a longer 'tail'.

Spontaneous remodeling: variation over time in the properties of individual synapses that do not result from specific activity histories.

Synaptic tenacity: the capacity of synapses to maintain their particular properties over behaviorally relevant timescales.

abolished (by eliminating the presynaptic proteins Munc13-1 and Munc13-2). These synaptic features were found to be similar to those observed in preparations from control animals, although postsynaptic currents were slightly reduced and less correlated with spine volume. In a second study [23], the near-complete suppression of presynaptic glutamate release in the mouse forebrain (by tetanus toxin expression) was found to have relatively modest effects on synapses and their properties, at least as judged by morphological measures. Although synapse numbers and spine densities were reduced in some brain regions (but not others), dendritic spines of all types formed at normal proportions.

These observations – that synapses of all sizes and types form at normal proportions – have profound implications: when synapses and dendritic spines are initially formed, they generally have small volumes and PSDs (e.g., [24–27]). Their subsequent conversion into large, mushroom-shaped spines is often thought to be driven by activity-dependent potentiation (reviewed in [28,29]; see also [30,31]). However, as mentioned above, large synapses, including mushroom-shaped spines, develop in normal proportions even when activity is essentially nonexistent [18,19,21,23]. It thus seems that activity-independent processes can generate the full repertoire of synaptic sizes [17], highlighting the potency of spontaneous synaptic remodeling.

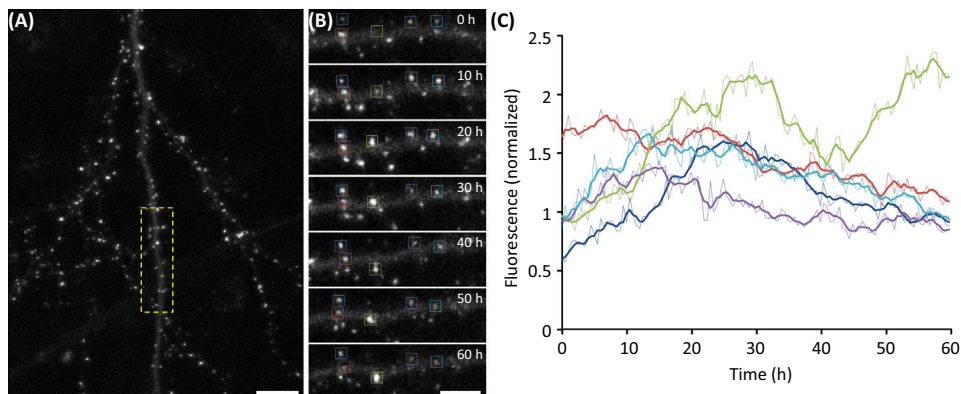
Spontaneous Synaptic Remodeling in Established Networks

Longitudinal observations of individual CNS synapses over many days in mature networks, both *in vitro* and *in vivo*, indicate that most synapses are persistent over these timescales, (e.g., [10,32] but see [9]; reviewed in [8]), although significant synapse formation and elimination are also observed (reviewed in [33–35]). Closer examination reveals, however, that properties of individual synapses, such as spine volume (e.g., [17,24,36–38]), presynaptic bouton volume [39,40], synaptic vesicle number [4,5,41], **active zone (AZ)** molecule content (Bassoon, Munc13-1) [4,42,43], and PSD protein content (PSD-95, Gria2, Gephyrin, GKAP, Shank, Zip45/Homer1c) [3,5,6,27,43–49] fluctuate considerably over these timescales (as illustrated for PSD-95 in Figure 1). Given the strong correlations between these measures and functional synaptic features such as synaptic vesicle release [42] and synaptic current amplitudes (reviewed in [8,29,50]), these fluctuations are likely to have functional consequences. Indeed, comparable fluctuations in connection strengths are observed when measured directly [51–53]. Furthermore, fluctuations tend to occur simultaneously in pre- and postsynaptic scaffolds of the same synapses [43], further supporting their functional significance.

As synapses in the aforementioned studies were embedded in active networks, their fluctuations reflect both spontaneous and activity-history-driven processes. Yet even when spontaneous activity is suppressed or eliminated altogether, fluctuations persist [3,6,17,40,54], although their characteristics can change (see below). These results suggest that in mature networks, as in developing networks, spontaneous synaptic remodeling is significant.

Relative Contributions of Specific Activity Histories and Spontaneous Processes to Synaptic Remodeling

The presence of significant activity-independent synaptic remodeling raises an important question: what are the relative contributions of specific activity histories and spontaneous processes to synaptic remodeling? This question might be addressed by comparing the magnitudes of changes they induce. As a rough estimate, changes to PSD sizes following experimental paradigms that induce long-term potentiation (measured using fluorescently tagged variants of PSD-95 and Homer1b) (e.g., [30,31]) seem to be of magnitude similar to



Trends in Neurosciences

Figure 1. Experimental Measurements of Synaptic Size Fluctuations. (A) A segment of a dendritic tree of a rat cortical neuron grown in culture expressing PSD-95 (a major glutamatergic PSD protein) tagged with GFP. Bar, 10 μm . (B) Images obtained at 10-h intervals of the segment enclosed in a broken rectangle in (A). Imaging was performed at 0.5-h intervals. Images shown here were created from maximum-intensity projections of ten sections at each time point. Bar, 5 μm . (C) 60-Hour traces of fluorescence intensities measured from the five synapses enclosed in color-coded squares in (B). Thin lines show original traces and thick lines show the same data after smoothing with a five-time-point kernel. Fluorescence values were normalized to average synapse fluorescence at $t = 0$. Images and data taken from the experiments described in [3].

that of baseline PSD size fluctuations observed *in vivo* using similar approaches [47,49]. However, these comparisons are made among different experimental systems and conditions (e.g., rat hippocampus CA1 pyramidal cells in cultured slices [30,31] and mouse layer 2/3 cortical pyramidal cells *in vivo* [47,49]).

The fluctuations observed in the latter studies represent the sum of both activity-dependent and -independent processes. As a step toward separating their respective contributions, the effects of suppressing activity were examined. Such experiments (in culture) [17,54] have shown that suppressing all spontaneous activity reduces the magnitude of glutamatergic synapse size fluctuations, but only by a factor of about two. Interestingly, little to no change was observed for GABAergic synapses [6,40].

A more direct estimate of the relative contributions is obtained by comparing pairs of synapses formed between the same axons and dendrites (see Figure 2 for an illustration of this approach). Such pairs share common activation histories that should drive similar remodeling in the two synapses. Conversely, size or remodeling differences between synapses belonging to the same pair would reflect spontaneous, activity-history-independent processes occurring autonomously at each synapse. This rationale was applied in several recent studies using electron microscopy reconstructions of hippocampal tissue [55], a fully reconstructed volume of mouse neocortex [56], and long-term imaging of cortical neurons in culture [54]. In general, it was found that such synapse pairs tend to be more similar than randomly chosen pairs. However, remodeling covariance over 48-h periods, as well as correlation coefficients of instantaneous sizes (spine volumes and PSD areas) were relatively small (~ 0.25 – 0.35). This led to an estimate that at most 40% of synaptic remodeling could be attributed to specific activity histories [54].

Collectively these findings suggest that the contributions of spontaneous processes and specific activity histories to synaptic remodeling are of similar magnitudes.

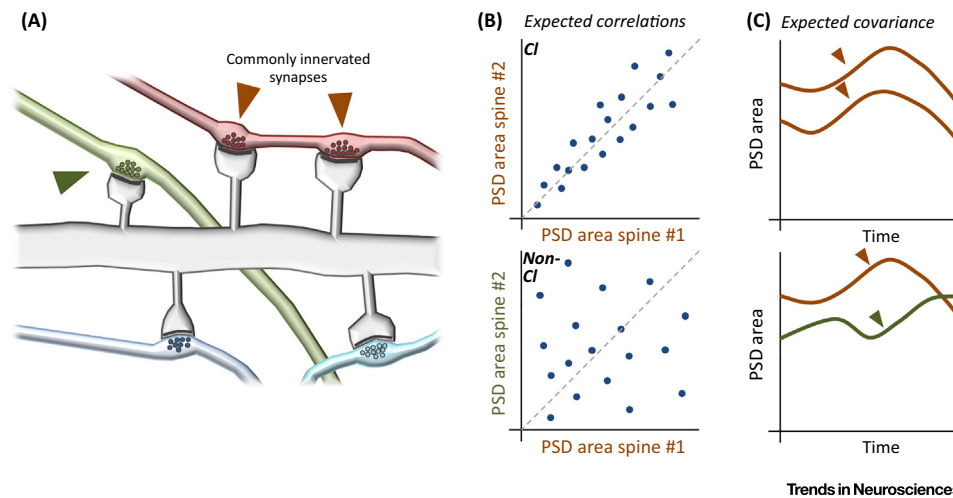


Figure 2. Pairs of synapses formed between the same axons and dendrites. (A) An illustration of a pair of commonly innervated (CI) synapses (red arrowheads); that is, synapses formed between the same axon (red) and the same dendrite (gray). Because of their shared axon and dendrite, such synapse pairs are assumed to have roughly similar activity histories. (B) Given their similar activity histories, the sizes of synapses belonging to the same CI synapse pair (top) would be expected to be more similar than the sizes of synapses innervated by different axons (non-CI; bottom), exemplified by arrowheads with differing colors in A. (C) Similarly, a higher degree of remodeling covariance might be expected between CI synapses (top) compared with a randomly chosen pair of synapses (bottom).

Characteristics, Sources, and Implications of Spontaneous Remodeling Processes

It appears that spontaneous remodeling is a central, inherent feature of CNS synapses, which merits in-depth analyses of its characteristics, sources, implications, and physiological importance. We mention a few of these here.

In the framework of synaptic plasticity, significance is attached to the specific configuration of synaptic strengths (synaptic weights) in a network. Therefore, it is important to characterize the rates at which spontaneous remodeling might ‘erode’ configurations of synaptic input strengths (Figure 3). Imaging studies in primary culture indicate that synaptic configurations erode significantly over timescales of a few days [3–6,27,38]. This becomes evident when synaptic size is plotted against initial size at increasingly greater time intervals, as illustrated in Figure 3B [6,27]. Not only are correlations reduced, but linear regression fits gradually become shallower while their offset terms grow larger. Suppressing network activity slows this process, but only by a factor of about two [3]. Interestingly, erosion rates of GABAergic synapse configurations seem to be insensitive to activity levels and slower than those of glutamatergic synapses in the same neurons [6].

Given enough time, will synaptic configurations erode completely? In studies published to date, monotonic erosion of synaptic configurations was reported [3–6,27,38]; however, over the timescales of such experiments, configurations did not erode entirely. Perhaps, given sufficient time, they would (Figure 3D, red broken lines). Alternatively however, they might conserve a semblance of their original configuration even after long times (Figure 3D, green broken lines). This could reflect limited mixing due to sizes fluctuating around synapse-specific set points (Figure 3E) defined, for example, by relatively stable core scaffolds [4,16]. Another possibility is that synaptic tenacity varies in the population such that the sizes of some synapses fluctuate much more than others. At present the extent of mixing and the heterogeneity of synaptic

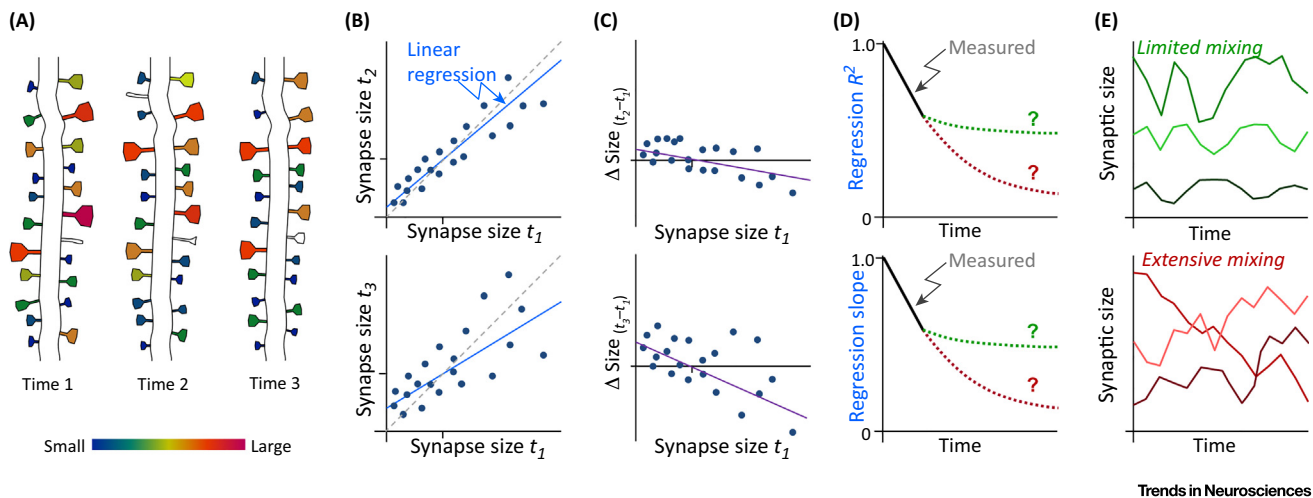


Figure 3. Erosion of Synaptic Configurations by Spontaneous Remodeling. (A) Illustration of a hypothetical dendritic segment at three consecutive time points. Sizes of individual spines are color coded according to the bar below. (B) Top: The size of each synapse at time t_2 is plotted against its size at an initial time t_1 . Light-blue lines are the linear regression fits whereas broken gray lines indicate the identity relationship. Bottom: Linear regression fits at a later time t_3 typically exhibit a lower goodness of fit (R^2) value and a shallower slope. (C) Changes in synaptic size as a function of initial size. Linear regression fits are shown as purple unbroken lines. Such lines cross the abscissa at the mean synaptic size, showing that, for synapses smaller than this mean, changes tend to be positive, whereas for synapses greater than this mean changes tend to be negative. For longer time intervals, the linear regression slopes gradually increase. (D) R^2 values (top) and slopes (bottom) of linear regression fits as in B decrease with time, providing a time scale for the erosion of synaptic configurations. What happens after very long times? This depends on the degree of mixing (E). If sizes fluctuate around synapse-specific set points [i.e., exhibit limited mixing (illustrated for three synapses; top)], R^2 values and slopes will settle at intermediate values [green broken lines in (D)]. If not (as illustrated for three synapses, bottom), they will diminish to zero [red broken lines in (D)]; at that time, all relationships with original synaptic configurations will be lost (i.e., these will have eroded entirely).

tenacity remain unknown. The rates at which synaptic configurations erode *in vivo* and how these might change with age [39] (see also [57]) or in association with various neurological disorders are not known either. These are crucial questions that call for very long measurements, particularly *in vivo* (see Outstanding Questions). Finally, the degree to which the observed erosion of synaptic configurations ultimately affects network function is unclear, but potential links between such erosion and memory decay [17] or declining mental abilities are plausible.

Although synaptic configurations deteriorate, the same analyses reveal other properties that emerge as stable invariants at the population level.

First, while single-synapse properties (e.g., spine volumes, PSD molecule contents) fluctuate, distributions of the same properties can be very stable [3,6,27,38]. Such distributions quantify the proportions of synapses of different sizes in the population; distribution stability implies that fluctuations in synaptic sizes are not merely diffusive (which would lead to the gradual broadening of size distributions) but reflect a dynamic equilibrium in which synaptic sizes are somehow constrained (Box 1). One way to appreciate the presence of such constraints is by plotting changes in synaptic sizes against their initial sizes at later times (Figure 3C). Such plots highlight the tendency of small synapses to grow larger and of large synapses to become smaller [3,6,27], with the magnitude such changes increasing with time.

Second, the shapes of distributions within synaptic populations (e.g., distributions of release probability, synaptic currents, spine volumes, PSD molecule and receptor contents) tend to be skewed; that is, to have a tail of particularly large or strong synapses (e.g., [6,18,19,27,58,59]; reviewed in [60]). It has been suggested that such distributions might be a consequence of

Box 1. Models of Synaptic Size Dynamics

How can stable, skewed size distributions arise in populations of synapses whose sizes fluctuate continually?

Suppose that all synapses start out having small sizes, and that their sizes change stochastically. At time $t + 1$ synaptic size is equal to its original size x_t plus some random value η_t ; that is, $x_{t+1} = x_t + \eta_t$ or, in terms of change at time t , $\Delta x_t = \eta_t$ [I]

In such diffusive fluctuations (random walk), distributions expand indefinitely (Figure 1A). Consequently, no stable distribution is ever reached (even if the average of η is zero).

The divergence of these distributions can be curtailed by adding a restraining 'force' $f(x_t)$ whose magnitude depends on x_t such that $\Delta x_t = f(x_t) + \eta_t$. In this formulation of constrained fluctuations (Langevin dynamics), changes are still random but a deterministic restraining force acts, in analogy to a spring, to maintain sizes near some mean value. In the simplest case, where the force is linear ($-\gamma x_t$) and fluctuations are Gaussian, the process becomes $\Delta x_t = \gamma x_t + \eta_t$ [II]

Such dynamics give rise to stable distributions (Figure 1B) but these are Gaussian, not skewed, implying that additional assumptions are necessary to recapitulate the skewed distributions observed physiologically.

Skewed distributions are often associated with multiplicative noise, meaning that fluctuation magnitudes depend on the variable's state (e.g., synaptic size). Multiplicative noise can be introduced through more complex formulations of x_{t+1} [17,38]; still, restraining forces are required to maintain stable distributions. In one study [17] good fits to experimental data were obtained by introducing restraining forces that differed for different synapse-size categories (as did state-dependent fluctuations). It is not obvious, however, how synapses would 'know' to which category they belong and when to switch the rules that govern their remodeling.

One particularly simple formulation, which bypasses the need for arbitrary categories, is the Kesten process. Here, multiplicative noise is built into the restraining force by supplementing it with a random component (ξ_t):

$$\Delta x_t = (-\gamma + \xi_t)x_t + \eta_t \quad \text{[III]}$$

This process entails two types of changes to synaptic size: continuous downscaling (on average), which includes state-dependent fluctuations, and an additive term that contains state-independent fluctuations (Figure 1C). The Kesten process was shown to describe faithfully many experimental observations [3,6,27,46], including: size fluctuations; stable, skewed distributions; growth of new synapses; and synaptic scaling (Box 2).

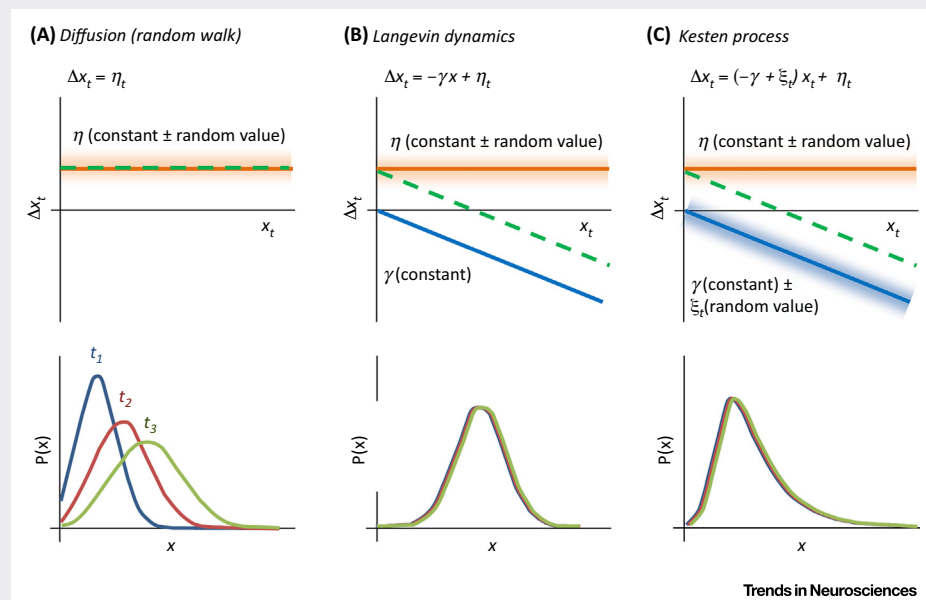


Figure 1. Shape and Stationarity of Synaptic Size Distributions Governed by Different Stochastic Processes. (A) Diffusion (random walk). (B) Langevin dynamics. (C) Kesten process. The top panels show the additive component (size independent; orange) and the restraining one (size dependent; blue) for each case, as well as their average sum (green). Bottom panels show the resulting distributions at three consecutive time points.

synaptic plasticity in several forms [61]. **Skewed distributions** emerge, however, even when activity is suppressed throughout network development [18,19], highlighting once again the significant contributions of spontaneous remodeling and its ability to generate full repertoires of synapses at correct proportions.

The realization that size fluctuations play crucial roles in synaptic remodeling has motivated several groups to explore this topic through computational modeling [17,27,29,38]. These models faithfully capture many of the experimental results and provide some insights into their potential origin. For example, they offer an explanation of how synaptic size fluctuations can give rise to skewed, stable size distributions (Box 1). They also provide an explanation for experimental findings showing that scaling of synaptic size distributions can occur without overt, uniform multiplicative scaling of individual synapse sizes [3,27] (Box 2).

A more recent computational study [62] has gone one step further, showing that all of these properties – size fluctuations and stable and skewed size distributions, as well as their scaling – can emerge naturally from stochastic assimilation and removal of synaptic molecules at synaptic sites, provided the two processes exhibit cooperativity. This model also recapitulates the internal spatial organization of synapses in the form of dynamic nanodomains (e.g., [63,64]). A more detailed model, where removal and aggregation are implemented by lateral diffusion of receptors bound to scaffold proteins, also provides good fits to measured distributions [65]. Since distribution properties can be described equally well by alternative mesoscopic models, further experimental and theoretical work is required to identify the key biophysical processes involved.

Concluding Remarks

The findings summarized above indicate that synaptic tenacity is inherently limited [3,4,6,43,54] or, using the terminology of Rumpel, Loewenstein, and others [34,35], that synapses are intrinsically ‘volatile’. How can the notions of synaptic plasticity and synaptic volatility be

Box 2. Synaptic Scaling as a Population-Level Phenomenon

Distributions of synaptic properties (e.g., synaptic current amplitude, glutamate receptor content, PSD size) are known to change following manipulations of spontaneous network activity levels, sensory input, and neuromodulatory tone. In particular, distributions were shown to scale (namely, retain their shape on a stretched/compressed x-axis) following such manipulations as illustrated in Figure 1. This scaling is often attributed to a global multiplicative change in the measured property such that all synapses formed on the same neuron change by some global scale factor [68]. This interpretation is particularly appealing as it suggests a mechanism for preserving the relative strengths of synaptic inputs, amounting to neuron-wide renormalization.

Most studies concerning this phenomenon were based on population-wide comparisons of synaptic properties before and after a manipulation. When individual synapses are tracked throughout the scaling process, however, additional information is revealed suggesting that **distribution scaling** can occur without overt, uniform multiplicative scaling of individual synapse sizes [3,27,46]. Indeed, these analyses directly show that the fold change differs markedly among synapses belonging to the same neuron. A mechanism for distribution scaling that is consistent with the individual-synapse dynamics entails changes in statistical parameters that govern spontaneous remodeling (Box 1). For example, such manipulations might change the slope of continuous downscaling (the average restraining force, $-\gamma$, in Box 1), the average magnitude of additive processes (the mean of parameter η in Box 1), or, possibly, the magnitude of multiplicative and additive fluctuations as illustrated in Figure 1A,C [3,6,27,46]. Therefore, synaptic scaling can emerge as a population-level phenomenon that does not necessarily preserve the relative strengths of synaptic inputs.

This result has important implications for relationships between activity and synaptic remodeling: it suggests that activity, in addition to driving remodeling through deterministic rules acting at the single-synapse level, can also modify steady-state properties of synaptic populations by parametrically affecting the features of stochastic processes that govern spontaneous remodeling.

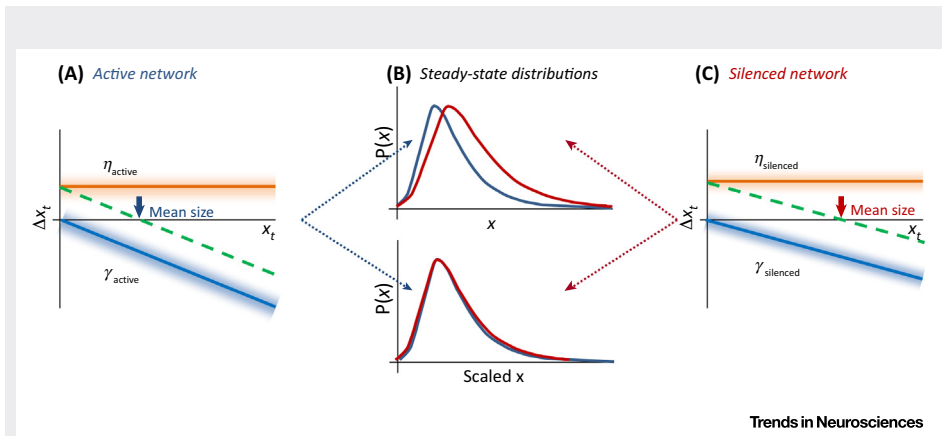


Figure 1. Scaling of Synaptic Size Distributions Can Be Driven by Parametric Changes in a Stochastic Process. (A,C) In this figure synaptic sizes are assumed to be governed by a stochastic Kesten process as in Box 1, resulting in stationary, skewed size distributions (B). Silencing the network slightly changes (weakens) the restraining force (the size-dependent component; blue) and/or increases the additive (size-independent) component (orange). This results in an increased mean synaptic size (the point at which the sum of the two components crosses the abscissa; arrowhead). Consequently, the distribution of synaptic sizes broadens [(B), top] but maintains its shape in scaled units [27] [(B), bottom].

reconciled? How can properties of individual neurons, such as sensory tuning and motor and place representation, as well as higher-level functions remain invariant if directed and spontaneous synaptic changes are, as suggested by various studies, of similar magnitude? Potential answers to these questions have been discussed in several recent reviews [33–35] and a subset of possible resolutions is summarized briefly in Box 3. Irrespective of interpretation, however, it is clear that the ideal notion of perfectly tenacious synapses is not supported by experimental findings (nor is it particularly plausible from a biological standpoint, given the

Box 3. Memory in the Presence of Limited Synaptic Tenacity

How can memory persist in the presence of significant spontaneous synaptic remodeling? This matter has been addressed in several recent reviews [33–35]. Some possible resolutions are listed briefly below.

- **Compound Connections:** Connections between neurons are often based on multiple synapses resulting in compound connections (reviewed in [69]). Consequently, fluctuations at the single-synapse level might average out at the compound connection level, improving the signal-to-noise ratio (SNR) of connection strengths and their plasticity.
- **Strength in numbers:** Persistent changes to network function might rely on the formation of new synapses and the elimination of others. The magnitude of spontaneous rewiring, however, seems to be comparable with directed rewiring ([70]; reviewed in [34,35]). An elegant model [71] tied these findings to the idea of compound connections mentioned above, suggesting that learning involves transitions between two relatively persistent states; namely, connections with few (zero or one) synapses and compound connections (approximately four or five synapses).
- **Massive redundancy:** Invariant function might involve vast numbers of neurons and synapses such that fluctuations at the single-synapse level become negligible. Indeed, the acquisition of a new motor skill in mice was estimated to involve ~4700 motor cortex neurons and ~410 000 synapses [72]. In this case, global properties other than specific patterns of synaptic weights might be the relevant system variables.
- **Diversity and selection:** Spontaneous remodeling might be functionally important for the generation of diverse, partially random synaptic configurations from which 'useful' configurations are thereafter selected; for example, through interactions with the environment (reviewed in [35]).
- **Synaptic sampling:** Spontaneous changes in synaptic properties might allow networks to explore and sample synaptic configurations for those most congruent with sensory input or desired functions and improve the ability of networks to generalize and compensate for unforeseen changes [73].
- **A paradigm shift?** The possibility remains that none of the potential explanations listed above will prove to be sufficient and that a paradigm shift will be ultimately required to understand the physiological substrate of learning and memory [66,67].

challenges to synaptic tenacity discussed above). Furthermore, spontaneous synaptic remodeling has important consequences that are not fully understood and are not always intuitive (see Outstanding Questions). When it comes to cognitive functions, long-term memory is one area where the notion of synaptic volatility raises perhaps some of the most challenging questions. In light of findings discussed in this Opinion article, and possibly others [34,66,67], age-old notions concerning relationships between histories of 'elementary brain-processes', connection strengths, and memory traces might need to be revisited; put differently (to paraphrase James [1]), modern science might need to improve on this explanation.

References

- James, W. (1890) Association. In *The Principles of Psychology* (Vol. 2), pp. 550–604, Henry Holt
- Hebb, D.O. (1949) *Organization of Behavior: A Neuropsychological Theory*, John Wiley and Sons
- Minerbi, A. et al. (2009) Long-term relationships between synaptic tenacity, synaptic remodeling, and network activity. *PLoS Biol.* 7, e1000136
- Fisher-Lavie, A. et al. (2011) Use dependence of presynaptic tenacity. *J. Neurosci.* 31, 16770–16780
- Zeidan, A. and Ziv, N.E. (2012) Neuroligin-1 loss is associated with reduced tenacity of excitatory synapses. *PLoS One* 7, e42314
- Rubinski, A. and Ziv, N.E. (2015) Remodeling and tenacity of inhibitory synapses: relationships with network activity and neighboring excitatory synapses. *PLoS Comput. Biol.* 11, e1004632
- Cohen, L.D. and Ziv, N.E. (2017) Recent insights on principles of synaptic protein degradation. *F1000Research* 6, 675
- Holtmaat, A. and Svoboda, K. (2009) Experience-dependent structural synaptic plasticity in the mammalian brain. *Nat. Rev. Neurosci.* 10, 647–658
- Attardo, A. et al. (2015) Impermanence of dendritic spines in live adult CA1 hippocampus. *Nature* 523, 592–596
- Qiao, Q. et al. (2016) Long-term stability of axonal boutons in the mouse barrel cortex. *Dev. Neurobiol.* 76, 252–621
- Berry, K.P. and Nedivi, E. (2017) Spine dynamics: are they all the same? *Neuron* 96, 43–55
- Bezprozvanny, I. and Hiesinger, P.R. (2013) The synaptic maintenance problem: membrane recycling, Ca^{2+} homeostasis and late onset degeneration. *Mol. Neurodegener.* 8, 23
- Wang, Y.C. et al. (2017) Presynaptic protein homeostasis and neuronal function. *Curr. Opin. Genet. Dev.* 44, 38–46
- Kosik, K.S. (2016) Life at low copy number: how dendrites manage with so few mRNAs. *Neuron* 92, 1168–1180
- Choquet, D. and Triller, A. (2013) The dynamic synapse. *Neuron* 80, 691–703
- Ziv, N.E. and Fisher-Lavie, A. (2014) Presynaptic and postsynaptic scaffolds: dynamics fast and slow. *Neuroscientist* 20, 439–452
- Yasumatsu, N. et al. (2008) Principles of long-term dynamics of dendritic spines. *J. Neurosci.* 28, 13592–13608
- Harms, K.G. and Craig, A.M. (2005) Synapse composition and organization following chronic activity blockade in cultured hippocampal neurons. *J. Comp. Neurol.* 490, 72–84
- Harms, K.J. et al. (2005) Synapse-specific regulation of AMPA receptor subunit composition by activity. *J. Neurosci.* 25, 6379–6388
- Ziv, N.E. and Garner, C.C. (2001) Principles of glutamatergic synapse formation: seeing the forest for the trees. *Curr. Opin. Neurobiol.* 11, 536–543
- Sigler, A. et al. (2017) Formation and maintenance of functional spines in the absence of presynaptic glutamate release. *Neuron* 94, 304–311
- Varoqueaux, F. et al. (2002) Total arrest of spontaneous and evoked synaptic transmission but normal synaptogenesis in the absence of Munc13-mediated vesicle priming. *Proc. Natl. Acad. Sci. U. S. A.* 99, 9037–9042
- Sando, R. et al. (2017) Assembly of excitatory synapses in the absence of glutamatergic neurotransmission. *Neuron* 94, 312–321
- Holtmaat, A. et al. (2006) Experience-dependent and cell-type-specific spine growth in the neocortex. *Nature* 441, 979–983
- Knott, G.W. et al. (2006) Spine growth precedes synapse formation in the adult neocortex *in vivo*. *Nat. Neurosci.* 9, 1117–1124
- Zito, K. et al. (2009) Rapid functional maturation of nascent dendritic spines. *Neuron* 61, 247–258
- Statman, A. et al. (2014) Synaptic size dynamics as an effectively stochastic process. *PLoS Comput. Biol.* 10, e1003846
- Bourne, J. and Harris, K.M. (2007) Do thin spines learn to be mushroom spines that remember? *Curr. Opin. Neurobiol.* 17, 381–386
- Kasai, H. et al. (2010) Structural dynamics of dendritic spines in memory and cognition. *Trends Neurosci.* 33, 121–129
- Meyer, D. et al. (2014) Balance and stability of synaptic structures during synaptic plasticity. *Neuron* 82, 430–443
- Bosch, M. et al. (2014) Structural and molecular remodeling of dendritic spine substructures during long-term potentiation. *Neuron* 82, 444–459
- Carrillo, J. et al. (2013) The long-term structural plasticity of cerebellar parallel fiber axons and its modulation by motor learning. *J. Neurosci.* 33, 8301–8307
- Clopath, C. et al. (2017) Variance and invariance of neuronal long-term representations. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 372, 20160161
- Mongillo, G. et al. (2017) Intrinsic volatility of synaptic connections – a challenge to the synaptic trace theory of memory. *Curr. Opin. Neurobiol.* 46, 7–13
- Chambers, A.R. and Rumpel, S. (2017) A stable brain from unstable components: emerging concepts and implications for neural computation. *Neuroscience* 357, 172–184
- Zuo, Y. et al. (2005) Development of long-term dendritic spine stability in diverse regions of cerebral cortex. *Neuron* 46, 181–189
- De Roo, M. et al. (2008) LTP promotes a selective long-term stabilization and clustering of dendritic spines. *PLoS Biol.* 6, e219
- Loewenstein, Y. et al. (2011) Multiplicative dynamics underlie the emergence of the log-normal distribution of spine sizes in the neocortex *in vivo*. *J. Neurosci.* 31, 9481–9488
- Grillo, F.W. et al. (2013) Increased axonal bouton dynamics in the aging mouse cortex. *Proc. Natl. Acad. Sci. U. S. A.* 110, E1514–E1523
- Schuemann, A. et al. (2013) Structural plasticity of GABAergic axons is regulated by network activity and GABA_A receptor activation. *Front. Neural Circuits* 7, 113
- Herzog, E. et al. (2011) *In vivo* imaging of intersynaptic vesicle exchange using VGLUT1 Venus knock-in mice. *J. Neurosci.* 31, 15544–15559
- Matz, J. et al. (2010) Rapid structural alterations of the active zone lead to sustained changes in neurotransmitter release. *Proc. Natl. Acad. Sci. U. S. A.* 107, 8836–8841

Outstanding Questions

Mixing and timescales: Given sufficiently long times, will individual synapses 'explore' the full range of synaptic sizes? Stated differently, to what degree do synapse sizes 'mix' (Figure 3D,E)? If mixing of synaptic sizes is substantial, over what timescales does such mixing occur? How do such timescales compare with the lifespans of individual synapses? Imaging individual synapses at high temporal resolution for extended timescales might provide clues about this critically important question.

Internal set points: If mixing of synaptic sizes is limited, this would indicate that synaptic sizes fluctuate around synapse-specific internal 'set points'. What might define such set points? How might these be modified?

Relationships between activity and synaptic size fluctuations: Are size fluctuations manifestations of specific activity histories and deterministic 'rules' or does activity parametrically affect stochastic processes that govern spontaneous synaptic remodeling (in analogy to temperature in chemical reactions and diffusive processes)? Given the strong influence that activity has on the dynamics of many synaptic molecules, the latter possibility cannot be ignored.

Spontaneous remodeling and network function: How does spontaneous synaptic remodeling affect the input–output relationships of individual neurons? How does it affect the fidelity of signal propagation within neuronal networks?

Biological sources: What gives rise to spontaneous synaptic remodeling? How is it related to biophysical dynamic processes and to internal synaptic structure, such as nanodomains and specific molecular assemblies?

Spontaneous remodeling and aging: How does spontaneous synaptic remodeling change with development, during aging, or in relation to neurological disorders or neurodegenerative conditions?

43. Fisher-Lavie, A. and Ziv, N.E. (2013) Matching dynamics of pre-synaptic and postsynaptic scaffolds. *J. Neurosci.* 33, 13094–13100
44. Marrs, G.S. *et al.* (2001) Rapid formation and remodeling of postsynaptic densities in developing dendrites. *Nat. Neurosci.* 4, 1006–1013
45. Kuriu, T. *et al.* (2006) Differential control of postsynaptic density scaffolds via actin-dependent and -independent mechanisms. *J. Neurosci.* 26, 7693–7706
46. Kaufman, M. *et al.* (2012) Long-term relationships between cholinergic tone, synchronous bursting and synaptic remodeling. *PLoS One* 7, e40980
47. Cane, M. *et al.* (2014) The relationship between PSD-95 clustering and spine stability *in vivo*. *J. Neurosci.* 34, 2075–2086
48. Pagès, S. *et al.* (2015) Single cell electroporation for longitudinal imaging of synaptic structure and function in the adult mouse neocortex *in vivo*. *Front. Neuroanat.* 9, 36
49. Gray, N.W. *et al.* (2006) Rapid redistribution of synaptic PSD-95 in the neocortex *in vivo*. *PLoS Biol.* 4, e370
50. Nishiyama, J. and Yasuda, R. (2015) Biochemical computation for spine structural plasticity. *Neuron* 87, 63–75
51. Tsukamoto-Yasui, M. *et al.* (2007) Active hippocampal networks undergo spontaneous synaptic modification. *PLoS One* 2, e1250
52. Usami, A. *et al.* (2008) Spontaneous plasticity of multineuronal activity patterns in activated hippocampal networks. *Neural Plast.* 2008, 108969
53. Loebel, A. *et al.* (2013) Matched pre- and post-synaptic changes underlie synaptic plasticity over long time scales. *J. Neurosci.* 33, 6257–6266
54. Dvorkin, R. and Ziv, N.E. (2016) Relative contributions of specific activity histories and spontaneous processes to size remodeling of glutamatergic synapses. *PLoS Biol.* 14, e1002572
55. Bartol, T.M. *et al.* (2015) Nanoconnectomic upper bound on the variability of synaptic plasticity. *Elife* 4, e10778
56. Kasthuri, N. *et al.* (2015) Saturated reconstruction of a volume of neocortex. *Cell* 162, 648–661
57. Mostany, R. *et al.* (2013) Altered synaptic dynamics during normal brain aging. *J. Neurosci.* 33, 4094–4104
58. Ariel, P. *et al.* (2013) Intrinsic variability in Pv, RRP, size, Ca²⁺ channel repertoire, and presynaptic potentiation in individual synaptic boutons. *Front. Synaptic Neurosci.* 4, 9
59. Zhang, Y. *et al.* (2015) Visualization of NMDA receptor-dependent AMPA receptor synaptic plasticity *in vivo*. *Nat. Neurosci.* 18, 402–407
60. Buzsáki, G. and Mizuseki, K. (2014) The log-dynamic brain: how skewed distributions affect network operations. *Nat. Rev. Neurosci.* 15, 264–278
61. Zheng, P. *et al.* (2013) Network self-organization explains the statistics and dynamics of synaptic connection strengths in cortex. *PLoS Comput. Biol.* 9, e1002848
62. Shomar, A. *et al.* (2017) Cooperative stochastic binding and unbinding explain synaptic size dynamics and statistics. *PLoS Comput. Biol.* 13, e1005668
63. Nair, D. *et al.* (2013) Super-resolution imaging reveals that AMPA receptors inside synapses are dynamically organized in nanodomains regulated by PSD95. *J. Neurosci.* 33, 13204–13224
64. MacGillavry, H.D. *et al.* (2013) Nanoscale scaffolding domains within the postsynaptic density concentrate synaptic AMPA receptors. *Neuron* 78, 615–622
65. Ranft, J. *et al.* (2017) An aggregation-removal model for the formation and size determination of post-synaptic scaffold domains. *PLoS Comput. Biol.* 13, e1005516
66. Trettenbrein, P.C. (2016) The demise of the synapse as the locus of memory: a looming paradigm shift? *Front. Syst. Neurosci.* 10, 88
67. Queenan, B.N. *et al.* (2017) On the research of time past: the hunt for the substrate of memory. *Ann. N. Y. Acad. Sci.* 1396, 108–125
68. Turrigiano, G. (2012) Homeostatic synaptic plasticity: local and global mechanisms for stabilizing neuronal function. *Cold Spring Harb. Perspect. Biol.* 4, a005736
69. Branco, T. and Staras, K. (2009) The probability of neurotransmitter release: variability and feedback control at single synapses. *Nat. Rev. Neurosci.* 10, 373–383
70. Nagaoka, A. *et al.* (2016) Abnormal intrinsic dynamics of dendritic spines in a fragile X syndrome mouse model *in vivo*. *Sci. Rep.* 6, 26651
71. Fauth, M. *et al.* (2015) Formation and maintenance of robust long-term information storage in the presence of synaptic turnover. *PLoS Comput. Biol.* 11, e1004684
72. Hayashi-Takagi, A. *et al.* (2015) Labelling and optical erasure of synaptic memory traces in the motor cortex. *Nature* 525, 333–338
73. Kappel, D. *et al.* (2015) Network plasticity as Bayesian inference. *PLoS Comput. Biol.* 11, e1004485